

*Low genetic variation and high differentiation across sky island populations of *Lupinus alopecuroides* (Fabaceae) in the northern Andes*

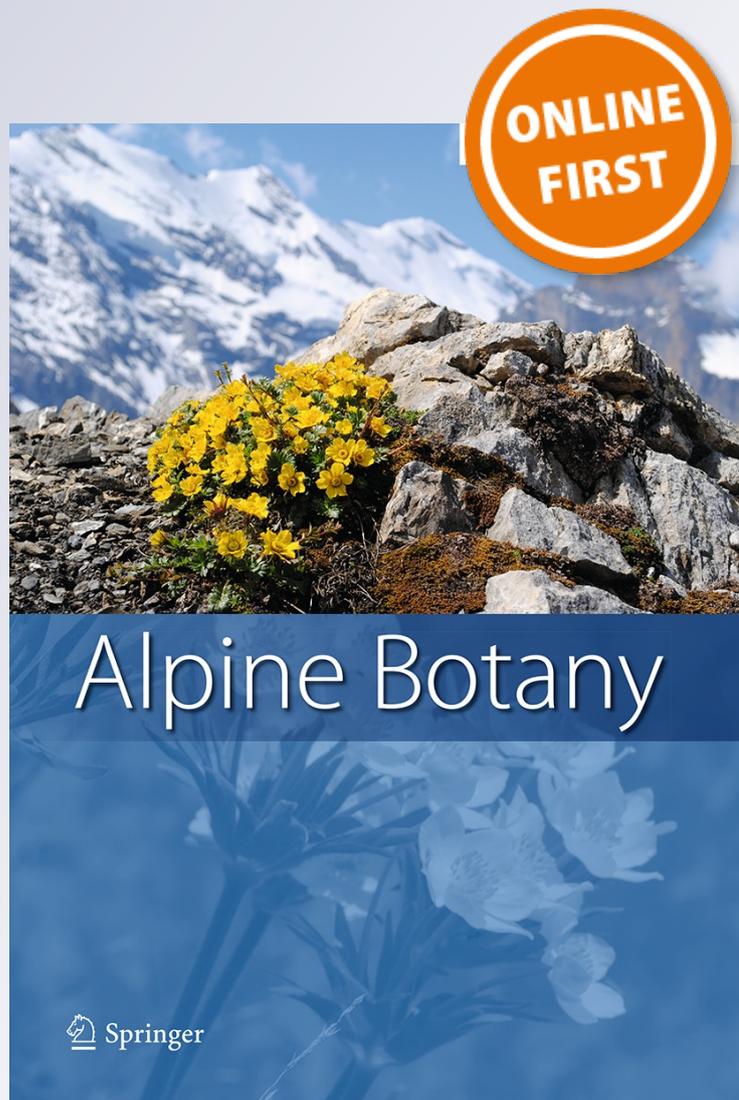
Diana L. A. Vásquez, Henrik Balslev, Michael Møller Hansen, Petr Sklenář & Katya Romoleroux

Alpine Botany

ISSN 1664-2201

Alp Botany

DOI 10.1007/s00035-016-0165-7



Your article is protected by copyright and all rights are held exclusively by Swiss Botanical Society. This e-offprint is for personal use only and shall not be self-archived in electronic repositories. If you wish to self-archive your article, please use the accepted manuscript version for posting on your own website. You may further deposit the accepted manuscript version in any repository, provided it is only made publicly available 12 months after official publication or later and provided acknowledgement is given to the original source of publication and a link is inserted to the published article on Springer's website. The link must be accompanied by the following text: "The final publication is available at link.springer.com".

Low genetic variation and high differentiation across sky island populations of *Lupinus alopecuroides* (Fabaceae) in the northern Andes

Diana L. A. Vásquez¹ · Henrik Balslev² · Michael Møller Hansen² · Petr Sklenář¹ · Katya Romoleroux³

Received: 18 November 2015 / Accepted: 11 April 2016
© Swiss Botanical Society 2016

Abstract The tropical alpine flora in the northern Andes has caught the attention of evolutionary biologists and conservationists because of the extent of its diversity and its vulnerability. Although population genetics studies are essential to understand how diversity arises and how it can be maintained, plant populations occurring above 4100 m a.s.l. in the so-called super-páramo have rarely been studied at the molecular level. Here, we use 11 microsatellite DNA markers to examine genetic structure in populations of *Lupinus alopecuroides*, a long-lived semelparous giant rosette known from only 10 geographically isolated populations. Each population is located on a different mountain top, of which three are in Colombia and seven in Ecuador. We analysed 220 individuals from all the ten known populations. We find low genetic variation in all but one of the populations. Four populations are completely monomorphic, and another five show only one polymorphic locus each. On the other hand, we find extremely high genetic differentiation between populations. We discuss the mechanisms that might cause this pattern, and we suggest that it is related to founder effects, lack of gene flow, and

autogamy. The genetic relationships among the populations, and the lack of correlation between the genetic and geographic distances also point to the importance of founder effects and colonization history in driving differentiation among the populations.

Keywords Population genetics · Genetic diversity · Founder effects · Páramo · Andean flora · *Lupinus alopecuroides*

Introduction

The highlands in the northern Andes host one of the World's most species-rich tropical alpine floras (Smith and Cleef 1988; Sklenář et al. 2011), characterized by rapid and recent evolution (van der Hammen et al. 1973; Hughes and Eastwood 2006; Madriñán et al. 2013). This is the so-called páramo ecosystem that lies between 3000 and 4900 m a.s.l., and the highest tiers of it, above 4100 m, is called the super-páramo (Cuatrecasas 1968; van der Hammen and Cleef 1986). Given its restricted geographic distribution, páramo plant diversity is severely threatened by habitat destruction due to global warming and land-use changes (Balslev and Luteyn 1992; Morueta-Holme et al. 2015; Vásquez et al. 2015). Although the páramo flora has received significant attention from evolutionary biologists and conservationists, population genetic studies are scarce (Oleas et al. 2012). To our knowledge, super-páramo plant populations have never been studied at the molecular level.

Patterns of genetic variation across populations are determined by various interrelated factors, and population history plays an important role (Ellstrand and Elam 1993; Hewitt 2000). In the northern Andes, Pleistocene climatic oscillations produced elevational range shifts that

Electronic supplementary material The online version of this article (doi:10.1007/s00035-016-0165-7) contains supplementary material, which is available to authorized users.

✉ Diana L. A. Vásquez
aparicio@natur.cuni.cz

¹ Department of Botany, Charles University, Benátská 2, 128 01 Prague 2, Czech Republic

² Department of Bioscience, Aarhus University, Ny Munkegade 116, Building 1540, 8000 Aarhus C, Denmark

³ Escuela de Ciencias Biológicas, Pontificia Universidad Católica del Ecuador, Av. 12 de Octubre y Roca Apdo. 2184, Quito, Ecuador

enormously impacted páramo species. During glaciations, species expanded across broader landscapes at lower elevations, establishing new populations and gene flow between populations that had been isolated. During interglacials, species were again restricted to geographically isolated “islands in the sky” (van der Hammen et al. 1973; Hooghiemstra et al. 2006; Graham 2009; Sklenář et al. 2011). Another historical factor that impacted the páramo populations is volcanism (Oleas et al. 2012). Volcanic activity is a driver of population extinction and recolonization (Carson et al. 1990; Beheregaray et al. 2003), and thus influences evolutionary trajectories of the populations.

We examined the genetic structure of populations of *Lupinus alopecuroides* Desr.—a long-lived semelparous giant rosette restricted to the super-páramo in Colombia and Ecuador. Semelparous giant rosettes are a distinctive feature of tropical alpine floras, and a fascinating example of convergent adaptive evolution to harsh high-elevation environments (Rundel et al. 1994; Sgorbati et al. 2004). Marcescent dead leaves of the rosette protect the meristem from nocturnal temperatures, which often are below zero (Monasterio 1986; Rundel et al. 1994), and lanate hairs on the up-to 1 m high inflorescence provide insulation for reproductive organs (Miller 1986) (Fig. 1). Semelparous plants maximize reproduction by investing all resources in a single reproductive episode because, due to harsh environmental conditions, future survival and reproduction are less likely (Young and Augspurger 1991).

The genetic structure of populations is also determined by species-specific biological traits such as dispersal ability and breeding system. One of the most common seed dispersal modes observed in lupines is ballistichory—a mechanical process in which the walls of the mature pod twist in opposite directions, firing the seeds 1–3 m away from the parent plant. Long-distance dispersal of lupine seeds is



Fig. 1 Typical habit and local distribution of *Lupinus alopecuroides* in the superpáramo of Sincholagua, Ecuador (Photo by Diana L. A. Vásquez)

usually afforded by waterways, human activities, and animals, mainly rodents (Milla and Iriondo 2011; Fagan et al. 2005; Maron and Kauffman 2006; Australian Department of Health and Ageing 2013). The reproductive biology of *L. alopecuroides* has remained unstudied. Other perennial species of *Lupinus* have asexual reproduction by vegetative regeneration, and there is no evidence of apomictic reproduction in the genus (Richards 1986). Most annual lupines are self-compatible while perennials are self-incompatible (Kittelson and Maron 2000). However, in the papilionoid legumes, to which *Lupinus* belongs, self-sterility is rare (East 1940; Kittelson and Maron 2000). For facultatively autogamous species, selfing may occur either as a result of insufficient pollinator visitation, or as a result of pollinator visits due to the close proximity of viable pollen to the receptive stigma (Pazy 1984; Karoly 1992), or due to transfer of pollen between flowers on the same individual (de Jong et al. 1993).

Another important factor influencing the genetic structure of natural populations is landscape (Slatkin 1987; Manel and Holderegger 2013). Populations of *Lupinus alopecuroides* are known from only ten geographically isolated mountain tops (Fig. 2). In geographically isolated populations, enhanced genetic drift and lack of gene flow result in genetic differentiation and loss of genetic diversity (Lesica and Allendorf 1995; Frankham et al. 2002; Allendorf and



Fig. 2 Geographic distribution of 10 locations (mountain tops), where populations of *Lupinus alopecuroides* were sampled. Map by Flemming Nørgaard

Luikart 2006). In these tropical “islands in the sky”, founder effects may also occur, as founders arrive over long distances from similar habitats, rather than from adjacent sites at lower elevations, which are ecologically different (Carson and Templeton 1984). Enhanced genetic drift, restricted gene flow, and founder events put the sky island populations of *L. alopecuroides* at risk of inbreeding depression, and potentially make them vulnerable to an extinction vortex (Frankham et al. 2002). In populations of *Puya raimondii*, which is another sky island long-lived semelparous giant rosette occurring around 4000 m a.s.l from Peru to northern Bolivia, low genetic variation was reported across 217 AFLP marker loci in 160 individuals from eight populations. Four populations were completely monomorphic, and each of the others displayed only one to three polymorphic loci. It was suggested that the causes of this pattern are high rates of selfing and repeated bottlenecks (Sgorbati et al. 2004). *Phaedranassa tunguraguae*, which is a long-lived, partially selfing plant, showed homozygote excess at 12 microsatellite loci. Its populations were restricted to a single valley in the Ecuadorian Andes, at 1500–2500 m a.s.l., and the most likely explanation suggested for this pattern was genetic drift due to small population size and restricted gene flow (Oleas et al. 2012).

It is well established that founding of a new population by a small number of individuals causes abrupt changes in allele frequencies that can lead to loss of genetic variation within populations and increased genetic differentiation among populations (Mayr 1954, Allendorf and Luikart 2006). However, founder effects are hard to detect, because it is difficult to disentangle their signatures from those of other evolutionary forces (Hoeck et al. 2010; Spurgin et al. 2014). In island populations of Berthelot's pipit (*Anthus berthelotii*), Spurgin et al. (2014) detected genetic and phenotypic changes that resulted from founder events, and suggested that, under lack of gene flow and selection, founder effects can persist for evolutionary time scales. In populations of silveryeyes (*Zosterops lateralis*), Clegg et al. (2002) demonstrated that sequential founder events produced divergence among populations. Finally, Prugnolle et al. (2005) have suggested that patterns of neutral genetic differentiation in human populations are the result of sequential founder events that occurred as small populations of modern humans migrated out of Africa.

Population genetics studies can help us understand how contemporary diversity arises and how it can be maintained (Frankham et al. 2002; Allendorf and Luikart 2006). Here, we use microsatellite markers to examine patterns of genetic variation in the sky island populations of *Lupinus alopecuroides*, and we discuss the factors underlying genetic variation and population structure. In particular, we address the following questions. (1) How high is genetic diversity within populations? (2) Are populations genetically

differentiated, and how is genetic variation distributed within and among the populations? (3) Is there gene flow between the populations? (4) How are the different populations related, and is there a geographical correlation?

Materials and methods

The species

Lupinus alopecuroides Desr. is a long-lived semelparous giant rosette known from only ten geographically isolated populations. Each population is located on a different mountain top, of which three are in Colombia and seven in Ecuador (Fig. 2). Within the populations, individuals are found forming dense clusters (Fig. 1). This might be related to the species' ballistic seed dispersal. We assume that long-distance dispersal of *L. alopecuroides* seeds, which weigh approx. 40 mg and measure 5×3 mm, is limited. Elsewhere rodents and human activity are important dispersers of *Lupinus* (Milla and Iriondo 2011; Fagan et al. 2005, Maron and Kauffman 2006; Australian Department of Health and Ageing 2013), but these agents are nearly absent in the super-páramo (Sklenář and Ramsay 2001, Rangel 2006). Anemochory or epizoochory are also unlikely in *L. alopecuroides*, since the seeds have a smooth surface without appendages or other adaptations that could facilitate external transport by animals. Some long-distance dispersal may be provided by waterways, white-tailed deer (*Odocoileus virginianus*), and domestic cattle, which have been seen to feed on *L. alopecuroides* pods (Vásquez, personal observation). The reproductive biology of the species has remained unstudied. Bumblebees are the most likely pollinator of *L. alopecuroides*, but their activity may be reduced by the harshness of the environment where this species grows (Dillon et al. 2006). During our fieldwork for this (10 weeks) and previous studies, we never observed insects visiting the flower, even on the sunniest days. We also observed that flowers remain closed for a long time after anthesis.

Population sampling

Exhaustive field and herbarium work in 2013–2014 permitted us to sample all ten known populations of *L. alopecuroides*, each of them located on a different mountain (Fig. 2). On the Cayambe and Sincholagua mountains, populations consist of three subpopulations clearly separated by 1–10 km (Table 1). Population size, elevation, and slope were recorded for each population and subpopulation (Table 1). Population size was estimated by counting all individuals or, when population size exceeded 50, by extrapolating it from the average number of individuals

Table 1 *Lupinus alopecuroides* populations and subpopulations with their respective population size, sample size, elevation, slope, area, mean allelic richness per loci, observed heterozygosity (Ho), expected heterozygosity (He), and number of polymorphic loci among the 11 analysed loci

Population	Subpopulation	Pop. Size	Sample size	Elevation (m a.s.l.)	Slope (°)	Area (m ²)	Mean allelic richness	Ho	He	No. polymorphic loci/pop
COCUY		~50	14	4150–4300	0–10	400,000	2.72	0.144	0.51	11
RUIZ		~400	22	4450–4600	20–30	35,000	1.08	0.01	0.04	1
PURACÉ		~600	23	4100–4200	20	22,000	1.08	0	0.04	1
COTACACHI		~60	19	4340–4350	30	700	1.11	0	0.036	1
IMBABURA		10	10	4350–4370	35	150	1	0	0	0
CAYAMBE	CAY1	~80	8	4468	0–20	22,500	1	0	0	0
	CAY2	~300	50	4425–4450	35	35,000				
	CAY3	9	9	4100–4200	40	300				
PICHINCHA		~250	18	4550–4600	20–30	38,000	1.09	0	0.04	1
ANTISANA		~50	12	4400–4450	35	600	1.16	0	0.06	1
SINCHOLAGUA	SINCH1	~100	19	4430–4438	30	5000	1	0	0	0
	SINCH2	~40	9	4430–4440	35	250				
	SINCH3	~100	11	4490–4495	40	4500				
CHIMBORAZO		8	8	4500–4600	10–20	300,000	1	0	0	0

Populations are sorted by latitude

counted in sub-plots. Population area was estimated from the polygon obtained from GPS-delimited boundaries using Google Earth Pro. Leaf tissue was randomly collected from 8 to 67 clearly distinct individuals, and stored in silica gel.

Molecular methods

DNA was extracted from leaf tissue of 220 individuals. Nine loci (Luna1, Luna3, Luna4, Luna6, Luna8, Luna12, Luna15, Luna17, Luna20) developed for *Lupinus nanus* (Molecular Ecology Resources Primer Development Consortium et al. 2012), and two loci (AG55-20-22, AG55-26-16) developed for *Lupinus microcarpus* (Drummond and Hamilton 2005) were amplified in four separate multiplexes, using QIAGEN Multiplex PCR Maxter Mix (Qiagen, Valencia, CA, USA). PCR products were electrophoresed on an automated capillary sequencer (3130xl Genetic Analyzer, Applied Biosystems, Foster City, CA, USA) with Genescan-600 (LIZ) size standard (Applied Biosystems). Sizes of alleles (in base pairs) were determined using GeneMarker (Soft Genetics).

Statistical analysis

To estimate within-population genetic diversity, allelic richness as well as observed heterozygosity (Ho), and unbiased expected heterozygosity (He) according to Nei (1987) were calculated for each population. Genetic differentiation among all populations was estimated using Weir and Cockerham's θ (1984), an estimator of F_{ST} , and R_{ST} , an

F_{ST} analogue based on the stepwise mutation model (Slatkin 1995). The program FSTAT 2.9.3 (Goudet 2001) was used for these analyses. Tests for significant deviation from Hardy–Weinberg equilibrium were conducted using exact tests implemented in Arlequin 3.5 (Excoffier and Lischer 2010). Genetic relationships among the populations were estimated using POPTREEW (Takezaki et al. 2014) to construct a neighbour-joining phenogram based on *dmu2* distances (Goldstein et al. 1995). Bootstrap values across loci were based on 10,000 permutations by locus. Spatial genetic structure was further explored by testing for isolation by distance using a Mantel test between a matrix of *dmu2* genetic distances (Goldstein et al. 1995) and a matrix of geographic distances as implemented in R package *ade4* (Jombart and Ahmed 2011).

Results

A total of 46 unambiguously scorable and reproducible alleles ranging from 75 to 346 bp were detected at 11 microsatellite loci across 220 individuals, representing 10 populations distributed throughout the species' geographic range. Numbers of alleles per locus ranged from 2 to 6. Monomorphic loci were observed in all the populations except COCUY, where all eleven loci were polymorphic. Five populations showed only one polymorphic locus each. The other four populations (CHIMBORAZO, IMBABURA, CAYAMBE, SINCHOLAGUA), were completely monomorphic (Table 1). Consequently, there was no

genetic variation between the subpopulations neither in CAYAMBE nor in SINCHOLAGUA. Moreover, CAYAMBE and SINCHOLAGUA were fixed for the same alleles (Online Resource 1). Therefore, all individuals within these populations were identical multilocus genotypes. Significant deviation from Hardy–Weinberg equilibrium was found across all non-monomorphic loci except Luna 3 (p value = 0.23) and Luna 17 (p value = 0.02) in the population COCUY. Genetic differentiation among all populations was high: R_{ST} = 0.65 and θ = 0.84 (95 % CI 0.753–0.900).

The neighbour-joining phenogram (Fig. 3) constructed from $dmu2$ distances (Goldstein et al. 1995) grouped all Ecuadorian populations except ANTISANA in a well-supported group (65 % bootstrap value). COTACACHI, CAYAMBE and SINCHOLAGUA were also a group with high bootstrap value (65 %). Test of isolation by distance yielded a non-significant outcome (p value = 0.416).

Discussion

Genetic diversity and genetic differentiation

Our analyses revealed extremely low genetic diversity in populations of *L. alopecuroides* throughout its geographic range. From the total of ten known populations, four were completely monomorphic (all individuals within the population were identical multilocus genotypes) and another five showed only one polymorphic locus each (Online Resource 1). On the other hand, the high θ and R_{ST} values show that the ten populations of *L. alopecuroides* are highly genetically differentiated. R_{ST} incorporates molecular distances between alleles, and if R_{ST} is substantially higher than θ , then this might indicate that mutations have contributed to differentiation, indicating a substantial phylogeographical signal (Slatkin 1995). In the present case, however, $\theta > R_{ST}$

suggesting that drift is the predominant force underlying differentiation. Our results also show that populations of *L. alopecuroides* are not in Hardy–Weinberg equilibrium. We suggest that this disequilibrium is due to selfing and enhanced genetic drift rather than null alleles, as it seems unlikely that null alleles should occur at all loci and populations, whereas selfing would be expected to affect all loci. Finally, we suggest that low genetic diversity within populations and high genetic differentiation between populations are related to founder effects, lack of gene flow, and/or autogamy.

Founder effects—in tropical sky islands, populations are likely founded by individuals arriving from long distances, rather than from the warm tropical surroundings (Carson and Templeton 1984). Long-distance dispersal of *L. alopecuroides* seeds might be limited since rodents and human activity, important dispersers of *Lupinus* elsewhere (Milla and Iriondo 2011; Fagan et al. 2005, Maron and Kauffman 2006; Australian Department of Health and Ageing 2013), are nearly absent in the super-páramo (Sklenář and Ramsay 2001, Rangel 2006). Therefore, *L. alopecuroides* populations are likely founded by a small number of individuals. This produces severe bottlenecks that cause loss of genetic variation and increases genetic differentiation (Mayr 1954). Furthermore, Pleistocene climatic oscillation and intensive volcanic activity could also promote founder and bottleneck events by causing population expansions and contractions, and by affecting the populations' extinction–recolonization rate (Carson and Templeton 1984; Carson et al. 1990; Beheregaray et al. 2003). Repeated founder events may have triggered genetic divergence among populations (Clegg et al. 2002), and loss of genetic variation through sequential bottlenecks, as has previously been invoked to explain low variation in other species (e.g. Hedrick 1996). Interestingly, COCUY is the only population that is not located on a volcanic mountain massif (Kroonenberg et al. 1990) and it showed the highest genetic diversity. However, further research is needed to test the influence of volcanic activity and Pleistocene climatic oscillations on within-population genetic variation.

Lack of gene flow—in isolated populations, genetic drift leads to fixation of random alleles, and hence to loss of genetic variation and high genetic differentiation due to the absence of gene flow (Lesica and Allendorf 1995; Frankham et al. 2002; Allendorf and Luikart 2006; Bech et al. 2009). The high θ value in our data indicates that populations of *L. alopecuroides* are fixed for different alleles, suggesting an absence of gene flow.

Autogamy—low within-population diversity and high differentiation among populations are often observed in selfing species (Hamrick and Godt 1996). In *L.*

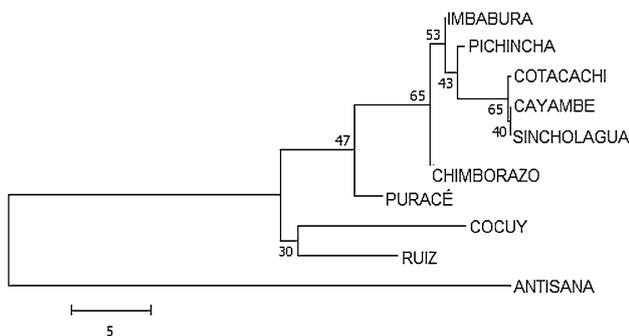


Fig. 3 Neighbour-joining dendrogram relating 10 *Lupinus alopecuroides* populations based on $dmu2$ distances (Goldstein et al. 1995). Bootstrap values across loci were based on 10,000 permutations by locus

alopecuroides, high rates of selfing may be expected due to low availability of pollinators (Vásquez personal observation). Autogamous self-fertilization, which is common among island plants (Carlquist 1974; Baker 1967), provides reproductive assurance when outcross pollination is limited by low availability of mates and/or pollinators (Karoly 1992). In the Venezuelan highlands, Berry and Calvo (1989) found near absence of pollinators and higher levels of selfing in four species of *Espeletia* (Asteraceae) growing above 4000 m a.s.l. Like lupines, *Espeletia* species are mainly pollinated by bumblebees. Significant decrease of diversity and activity levels of pollinators with elevation was also found in the Chilean Andes (Arroyo et al. 1985). Although, high inbreeding was indicated in the COCUY population and self-sterility is rare among lupines (East 1940; Kittelson and Maron 2000), further research is needed to demonstrate autogamy in *L. alopecuroides* and its role in determining the population's genetic structure.

Genetic relationships between populations

The neighbour-joining phenogram (Fig. 3) identified a well-supported group consisting of all Ecuadorian populations except ANTISANA. Within that group, CAYAMBE and SINCHOLAGUA, which were shown to be genetically indistinguishable (Online Resource 1), are the only populations located on the eastern Ecuadorian Cordillera. The population ANTISANA was shown to differ genetically from the rest of the populations. Interestingly, this population was also phenotypically different from the rest. In comparison with other populations, individuals have smaller leaves and inflorescence, and the overall size of the plants was also smaller (Vásquez, personal observation).

Finally, correlation between genetic and geographic distances of pair of populations was not significant, as was shown by the test for isolation by distance. This indicates that the genetic structure of *L. alopecuroides* populations is not the simple product of geographic spatial structure, pointing towards the importance of colonization history and founder effects in driving differentiation among populations. Although it has been heavily debated, the role of founder effects in evolution remains poorly understood (Clegg et al. 2002; Spurgin et al. 2014). It is unclear if founder effects can persist through evolutionary time when at the same time ongoing selection, mutation, gene flow and drift affect the genetic composition of populations (Hoeck et al. 2010; Spurgin et al. 2014). In this regard, it has been suggested that, depending on the severity and the continuity of the founder effects (Clegg et al. 2002), and on the extent of gene flow and selection, founder effects may be detectable in present populations, and may play an important role in the initial stages of speciation (Spurgin et al. 2014).

Possible effects of microsatellite cross-species amplification

Success of cross-species transfer of microsatellite markers depends on the evolutionary distance between the source and the target species (Rossetto 2001; Selkoe and Toonen 2006; Barbara et al. 2007). *Lupinus alopecuroides*, *L. microcarpus*, and *L. nanus* belong to a large western New world “super-radiation” (5.0–13.2 Ma) that comprises the western lupines from North America, Mexico and the Andes (Drummond et al. 2012). Low sequence divergence in adaptive radiations allows transfer of polymorphic microsatellite markers between species of the same sub-family and beyond (Barbara et al. 2007; Bezault et al. 2012). Otherwise, in plants, cross-species transfer of polymorphic markers is likely to be successful mainly within genera (Rossetto 2001; Barbara et al. 2007).

Cross-species transfer of microsatellites markers is often accompanied by a decrease in allelic diversity (Selkoe and Toonen 2006). However, if cross-species amplification underlies the low variation observed in the present study, then this should be a species-wide effect, whereas low variation within populations but higher variation in the species as a whole would suggest that low variation reflects demographic history. The latter was the case in our study; the total number of alleles for the species ranged from 2 to 6 across loci (Online Resource 1). One population (COCUY) in fact exhibited most of the observed alleles, whereas the other populations were mostly fixed for different alleles. We, therefore, conclude that the observed patterns of microsatellite DNA variation reflect genuine demographic processes and are not a result of reduced variation due to cross-species transfer.

Conclusions

Our study reveals extremely low genetic diversity within populations and high genetic differentiation between populations of *L. alopecuroides* across its range. We suggest that this pattern is related to founder effects, lack of gene flow, and possibly autogamy. However, further research is needed to provide evidence for autogamy in *L. alopecuroides*. The genetic relationships between the populations and the lack of correlation between genetic and geographic distances point to the importance of colonization history and founder effects in determining the populations' genetic structure. Although based on a limited number of markers, our study gives insights into the evolution of the unexplored but fascinating sky island Andean flora. Moreover, *L. alopecuroides* provides an exceptional opportunity for understanding the role of founder effects in evolution. Future studies should focus on reconstructing the colonization history of *L.*

alopecuroides, including the impact of volcanism and Pleistocene climatic oscillations on their population dynamics. Moreover, the low genetic variation and supposedly low adaptive potential in most populations suggest that the species could be particularly susceptible to anthropogenic disturbance. Formulation of a conservation strategy to protect *L. alopecuroides* is, therefore, strongly recommended.

Acknowledgments Thanks to Eva Dušková and Lenka Flašková for laboratory assistance, and to Jan Brotánek and Vojtěch Zeisek for help with fieldwork. We also thank Filip Kolář for helpful discussions and the Fond Mobility of Charles University in Prague for financial support.

Compliance with ethical standards

Conflict of interest The authors declare that they have no conflict of interest.

References

- Allendorf FW, Luikart G (2006) Conservation and the genetics of populations. Blackwell, London
- Arroyo MTK, Armesto JJ, Primack RB (1985) Community studies in pollination ecology in the high temperate Andes of central Chile II. Effect of temperature on visitation rates and pollination possibilities. *Plant Syst Evol* 149:187–203
- Australian Government. Department of Health and Ageing, Office of the Gene Technology Regulator (2013) The biology of *Lupinus L.* (lupin or lupine). [http://ogtr.gov.au/internet/ogtr/publishing.nsf/Content/biologylupin2013-toc/\\$FILE/biology-lupin20132.pdf](http://ogtr.gov.au/internet/ogtr/publishing.nsf/Content/biologylupin2013-toc/$FILE/biology-lupin20132.pdf). Accessed 5 March 2016
- Baker HG (1967) Support for Baker's law—as a rule. *Evolution* 21:853–856
- Balslev H, Luteyn JL (1992) Páramo: an Andean ecosystem under human influence. Academic Press, London
- Barbara T, Palma-Silva C, Paggi GM, Bered F, Fay MF, Lexer C (2007) Cross-species transfer of nuclear microsatellite markers: potential and limitations. *Mol Ecol* 16:3759–3767
- Bech N, Boissier J, Drovetski S, Novoa C (2009) Population genetic structure of rock ptarmigan in the 'sky islands' of French Pyrenees: implications for conservation. *Anim Conserv* 12:138–146
- Beheregaray LB, Ciofi C, Geist D, Gibbs JP, Caccone A, Powell JR (2003) Genes record a prehistoric volcano eruption in the Galápagos. *Science* 302:75
- Berry PE, Calvo RN (1989) Wind pollination, self-incompatibility, and altitudinal shifts in pollination systems in the high Andean genus *Espeletia* (Asteraceae). *Am J Bot* 76:1602–1614
- Bezault E, Rognon X, Gharbi K, Baroiller JF, Chevassus B (2012) Microsatellites cross-species amplification across some African cichlids. *Int J Evolut Biol*. doi:10.1155/2012/870935
- Carlquist S (1974) Island biology. Columbia University Press, New York
- Carson HL, Templeton AR (1984) Genetic revolutions in relation to speciation phenomena: the founding of new populations. *Annu Rev Ecol Syst* 15:97–131
- Carson HL, Lockwood JP, Craddock EM (1990) Extinction and recolonization of local populations on a growing shield volcano. *Proc Natl Acad Sci USA* 87:7055–7057
- Clegg SM, Degnan SM, Kikkawa J, Moritz C, Estoup A, Owens IP (2002) Genetic consequences of sequential founder events by an island-colonizing bird. *Proc Natl Acad Sci USA* 99:8127–8132
- Cuatrecasas J (1968) Páramo vegetation and its life-forms. In: Troll C (ed) Geoeology of mountainous regions of the tropical America. Colloquium geographicum 9. Geographisches Institut, Bonn, pp 163–186
- de Jong TJ, Waser NM, Klinkhamer PGL (1993) Geitonogamy: the neglected side of selfing. *Trends Ecol Evol* 8:321–325
- Dillon ME, Frazier MR, Dudley R (2006) Into thin air: physiology and evolution of alpine insects. *Integr Comp Biol* 46:49–61
- Drummond CS, Hamilton MB (2005) Isolation and characterization of nuclear microsatelliteloci in *Lupinus* group *Microcarpi* (Leguminosae). *Mol Ecol Notes* 5:510–513
- Drummond CS, Eastwood RJ, Miotto ST, Hughes CE (2012) Multiple continental radiations and correlates of diversification in *Lupinus* (Leguminosae): testing for key innovation with incomplete taxon sampling. *Syst Biol* 61:443–460
- East EM (1940) The distribution of self-sterility in the flowering plants. *Proc Am Philos Soc* 82:449–518
- Ellstrand NC, Elam DR (1993) Population genetic consequences of small population size: implications for plant conservation. *Annu Rev Ecol Syst* 24:217–242
- Excoffier L, Lischer HEL (2010) Arlequin suite ver 3.5: a new series of programs to perform population genetics analyses under Linux and Windows. *Mol Ecol Resour* 10:564–567
- Fagan WF, Lewis M, Neubert MG, Aumann C, Apple JL, Bishop JG (2005) When can herbivores slow or reverse the spread of an invading plant? A test case from Mount St. Helens. *Am Nat* 166:669–685
- Frankham R, Ballou JD, Briscoe DA (2002) Introduction to conservation genetics. Cambridge University Press, Cambridge
- Goldstein DB, Ruiz Linares A, Cavalli-Sforza LL, Feldman MW (1995) Genetic absolute dating based on microsatellites and the origin of modern humans. *Proc Natl Acad Sci USA* 92:6723–6727
- Goudet J (2001) FSTAT, a program to estimate and test gene diversities and fixation indices (version 2.9.3)
- Graham A (2009) The Andes: a geological overview from a biological perspective. *Ann Missouri Bot Gard* 96:371–385
- Hamrick JL, Godt MJW (1996) Effects of life history traits on genetic diversity in plant species. *Philos Trans R Soc B* 351:1291–1298
- Hedrick PW (1996) Bottleneck(s) or metapopulation in cheetahs. *Conserv Biol* 10:897–899
- Hewitt GM (2000) The genetic legacy of the Quaternary ice ages. *Nature* 405:907–913
- Hoek PE, Bollmer JL, Parker PG, Keller LF (2010) Differentiation with drift: a spatio-temporal genetic analysis of Galápagos mockingbird populations (*Mimus* spp.). *Philos Trans R Soc B* 365:1127–1138
- Hooghiemstra H, Wijninga VM, Cleef AM (2006) The paleobotanical record of Colombia: implications for biogeography and biodiversity. *Ann Missouri Bot Gard* 93:297–324
- Hughes C, Eastwood R (2006) Island radiation on a continental scale: exceptional rates of plant diversification after uplift of the Andes. *Proc Natl Acad Sci USA* 103:10334–10339
- Jombart T, Ahmed I (2011) adegenet 1.3-1: new tools for the analysis of genome-wide SNP data. *Bioinformatics* 27:3070–3071
- Karoly K (1992) Pollinator limitation in the facultatively autogamous annual, *Lupinus nanus* (Leguminosae). *Am J Bot* 79:49–56
- Kittelson PM, Maron JL (2000) Outcrossing rate and inbreeding depression in the perennial yellow bush lupine, *Lupinus arboreus* (Fabaceae). *Am J Bot* 87:652–660
- Kroonenberg SB, Bakker JG, van der Wiel AM (1990) Late Cenozoic uplift and paleogeography of the Colombian Andes: constraints on the development of high-Andean biota. *Geol Mijnbouw* 6:279–290
- Lesica P, Allendorf FW (1995) When peripheral populations are valuable for conservation. *Conserv Biol* 9:753–760

- Madriñán S, Cortés AJ, Richardson JE (2013) Páramo is the world's fastest evolving and coolest biodiversity hotspot. *Front Genet* 4:192
- Manel S, Holderegger R (2013) Ten years of landscape genetics. *Trends Ecol Evol* 28:614–621
- Maron JL, Kauffman MJ (2006) Habitat-specific impacts of multiple consumers on plant population dynamics. *Ecology* 87:113–124
- Mayr E (1954) Change of genetic environment and evolution. In: Huxley J, Hardy AC, Ford EB (eds) *Evolution as a process*. Allen and Unwin, London, pp 157–180
- Milla R, Iriondo AEM (2011) Congruence between geographic range distribution and local competitive ability of two *Lupinus* species. *Am J Bot* 98:1456–1464
- Miller GA (1986) Pubescence, floral temperature and fecundity in species of *Puya* (Bromeliaceae) in the Ecuadorian Andes. *Oecologia* 70:1432–1939
- Molecular Ecology Resources Primer Development Consortium, Abreu AG, Albaina A, Alpermann TJ, Apkenas VE et al (2012) Permanent genetic resources added to molecular ecology resources database 1 October 2011–30 November 2011. *Mol Ecol Resour* 12:374–376
- Monasterio M (1986) Adaptive strategies of Espeletia in the Andean desert páramo. In: Vuilleumier F, Monasterio M (eds) *High altitude tropical biogeography*. Oxford University Press, New York, pp 49–80
- Morueta-Holme N, Engemann K, Sandoval-Acuña P, Jonas JD, Segnitz RM, Svenning JC (2015) Strong upslope shifts in Chimborazo's vegetation over two centuries since Humboldt. *Proc Natl Acad Sci USA* 112:12741–12745
- Nei M (1987) *Molecular evolutionary genetics*. Columbia University Press, New York
- Oleas NH, Meerow AW, Francisco-Ortega J (2012) Population dynamics of the endangered plant, *Phaedranassa tunguraguae*, from the Tropical Andean Hotspot. *J Hered* 103:557–569
- Pazy B (1984) Insect induced self-pollination. *Plant Syst Evol* 144:315–320
- Prugnolle F, Manica A, Balloux F (2005) Geography predicts neutral genetic diversity of human populations. *Curr Biol* 15:R159–R160
- Rangel O (2006) The biodiversity of the Colombian páramo and its relation to antropoghenic impact. In: Spehn EM et al (eds) *Land use change and mountain biodiversity*. CRC Press, Boca Raton, pp 108–109
- Richards AJ (1986) *Plant breeding systems*. George Allen & Unwin, London
- Rossetto M (2001) Sourcing of SSR markers from related plant species. In: Henry RJ (ed) *Plant genotyping: the DNA fingerprinting of plants*. CAB International, Oxford, pp 211–224
- Rundel PW, Smith AP, Meinzer FC (1994) *Tropical alpine environments: plant form and function*. Cambridge University Press, Cambridge
- Selkoe KA, Toonen RJ (2006) *Microsatellites for ecologists: a practical guide to using and evaluating microsatellite markers*. *Ecol Lett* 9:615–629
- Sgorbati S, Labra M, Grugni E et al (2004) A survey of genetic diversity and reproductive biology of *Puya raimondii* (Bromeliaceae), the endangered queen of the Andes. *Plant Biol* 6:222–230
- Sklenar P, Ramsay PM (2001) Diversity of zonal páramo plant communities in Ecuador. *Divers Distrib* 7:113–124
- Sklenář P, Duškova E, Balslev H (2011) Tropical and temperate: evolutionary history of páramo flora. *Bot Rev* 2:71–108
- Slatkin M (1987) Gene flow and the geographic structure of natural populations. *Science* 236:787–792
- Slatkin M (1995) A measure of population subdivision based on microsatellite allele frequencies. *Genetics* 139:457–462
- Smith JMB, Cleef AM (1988) Composition and origins of the world's tropicalpine floras. *J Biogeogr* 15:631–645
- Spurgin LG, Illera JC, Jorgensen TH, Dawson DA, Richardson DS (2014) Genetic and phenotypic divergence in an island bird: isolation by distance, by colonization or by adaptation? *Mol Ecol* 5:1028–1039
- Takezaki N, Nei M, Tamura K (2014) POPTREEW : web version of POPTREE for constructing population trees from allele frequency data and computing other population statistics. *Mol Biol Evol* 31:1622–1624
- van der Hammen T, Cleef AM (1986) Development of the high Andean páramo flora and vegetation. In: Vuilleumier F, Monasterio M (eds) *High altitude tropical biogeography*. Oxford University Press, Oxford, pp 153–201
- van der Hammen T, Werner JH, van Dommelen H (1973) Palynological record of the upheaval of the Northern Andes: a study of the Pliocene and Lower Quaternary of the Colombian Eastern Cordillera and the early evolution of its high-Andean biota. *Palaeogeogr Palaeoclimatol* 16:1–24
- Vásquez DLA, Balslev H, Sklenář P (2015) Human impact on tropical-alpine plant diversity in the northern Andes. *Biodivers Conserv* 24:2673–2683
- Weir BS, Cockerham CC (1984) Estimating F-statistics for the analysis of population structure. *Evolution* 38:1358–1370
- Young TP, Augspurger CK (1991) Ecology and evolution of long-lived semelparous plants. *Trends Ecol Evol* 6:285